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**Bone Regeneration in femur defects
in rabbits treated with a e-PTFE
and a VBR titanium membrane**

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01 Poster Bone Regeneration VBR



Bone regeneration in femur defects in rabbits treated with a e-PTFE and a VBR titanium membrane

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INTRODUCTION

The need to insert dental implants with a guided prosthetic axis in patients with atrophy of the jaw-bone has determined the use of numerous techniques of GUIDED BONE REGENERATION or AUGMENT. GBR/GBA (1,2,3). The guided bone regeneration is based on the use of a membrane that, acting as a mechanical semi-waterproof barrier, excludes the connective and epithelial cells from the surgical repair and, at the same time, favours the osteo-promoting cells invasion (4).

The fundamental conditions to obtain the bone regeneration are:

maximum contact surface between the surrounding bone and the coagulum (5,6)

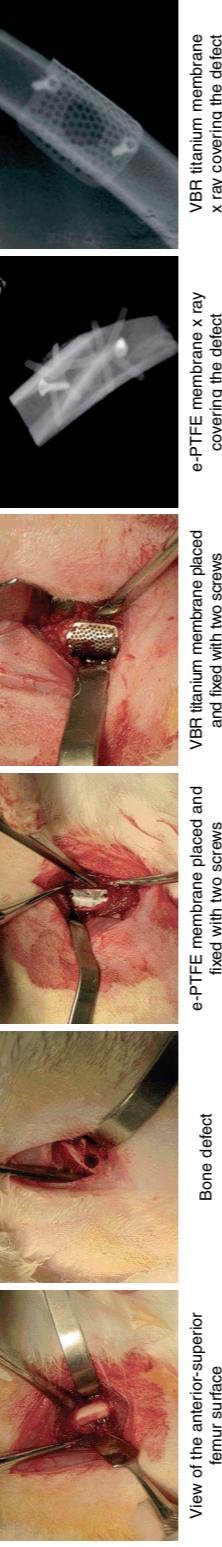
maximum overlying soft tissues respect

In bone defects with low space making capacity (open defects) the membrane, as well as creating a protective and semi-waterproof compartment for the coagulum, keeps the space (space making effect) favouring the osteogenesis process (7).

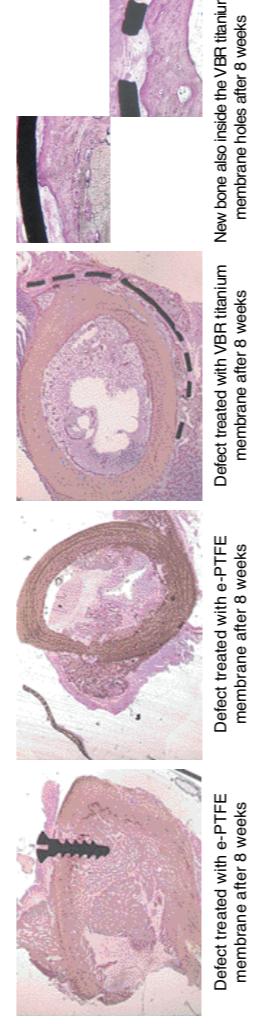
AIM

The aim of this work was an histological study of the bone healing of rabbits treated with an e-PTFE titanium reinforced membrane (W. L. Gore & Associates, Flagstaff, AZ, USA) and a new titanium membrane VBR (Valve Bone Regeneration, Oniplant, Cordenons, Italy) and their space making capacity.

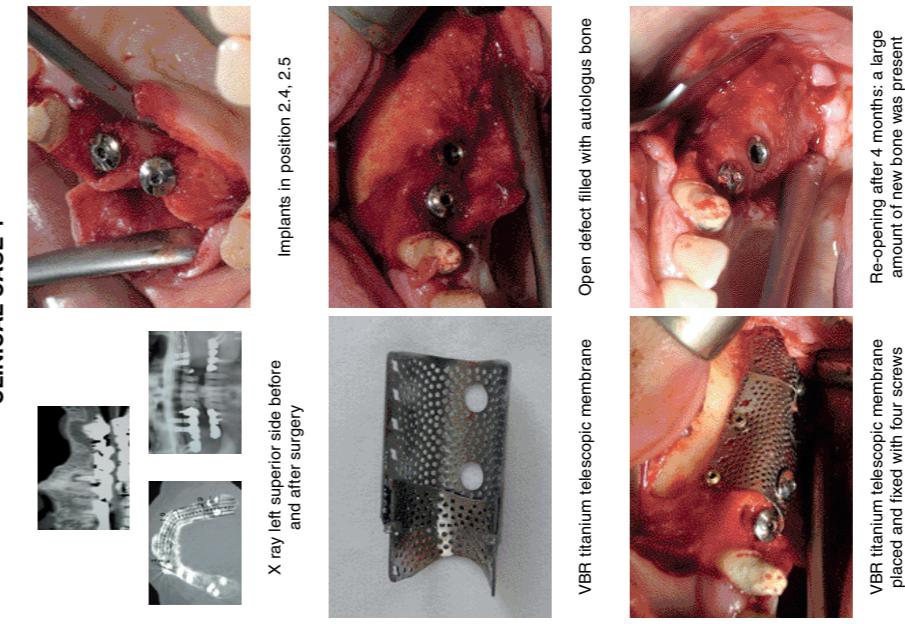
SURGICAL PROCEDURE



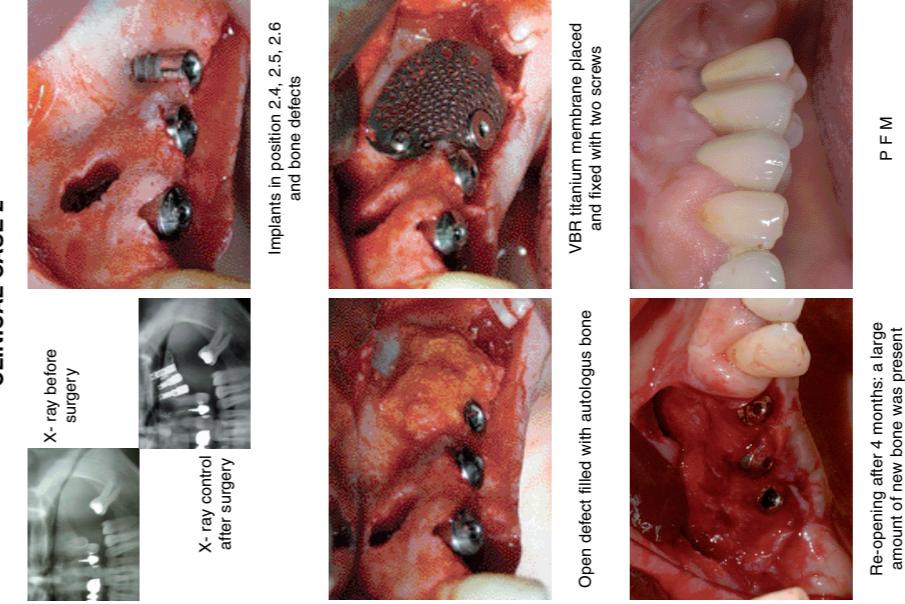
RESULTS



CLINICAL CASE 1

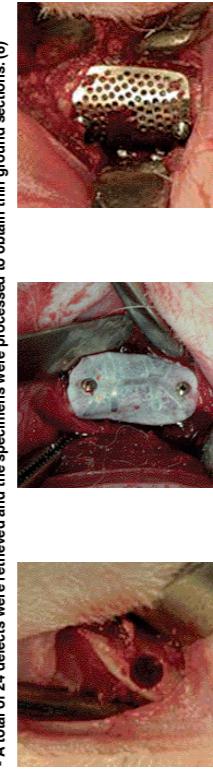


CLINICAL CASE 2



MATERIAL AND METHODS

- Twelve New Zealand rabbits, weighing about 2.5 Kg, were used.
- One defect (6mm x 6mm) was created in each femur.
- Twelve defects were covered with e-PTFE membranes (control defects).
- Twelve defects were covered with VBR titanium membranes (test defects).
- The rabbits were killed after 8 weeks and the block sections, containing the bone defects, were retrieved.
- A total of 24 defects were retrieved and the specimens were processed to obtain thin ground sections (8).



- e-PTFE membrane x ray covering the defect
- VBR titanium membrane x ray covering the defect

CONCLUSIONS

- All the defects of both groups were completely filled by mature, lamellar bone.
- No inflammatory cells infiltrate was present. No multi-nucleated giant cells were present.
- Newly formed bone was in close direct contact with both membranes and no gaps were present. Both membranes adhered closely to the bone defects.
- The e-PTFE membrane appeared to be compressed, in a few areas, by the overlying soft tissues.
- The VBR membrane did not appear compressed in any areas.
- No differences were found in the quantity of the bone regeneration using these two types of membranes and both membranes have shown a high degree of biocompatibility, and did not induce any ward effects. (9,10,11,12,13)

BIBLIOGRAPHY

1. Hurley AL, Stinchfield FE, Bassett CAL, Lyon WH. The role of soft tissue in osteogenesis (abstract). J Bone Joint Surg 1959;41A:1243.
2. Dahlin C, Sennérby L, Lekom U et al. Generation of new bone around titanium implants using a membrane technique: an experimental study in rabbits. Int J Oral Maxillofac Implants 1989;4:1925.
3. Dahlin C, Linde A, Gottlow J, Nyman S. Healing of bone defects by guided bone regeneration. Plast Reconstr Surg 1988;81:672-676.
4. Jovanovic SA, Schenk RK, Ossini M et al. Subperiosteal bone formation around dental implants: an experimental dog study. Int J Oral Maxillofac Impl 1995;10:23-31.
5. Weigel PH, Fuller GM, Leboeuf RD. A model for the role of hyaluronic acid and fibrin in the early events during the inflammatory response and wound healing. J Theor Biol 1986;119:219-34.
6. Bostrom MPG. Expression of bone morphogenetic proteins in fracture healing. Clin Orthop 1998;355:S116-S123.
7. Schenck RK, Buser D, Hardwick WR et al. Healing pattern of bone regeneration in membrane-protected defects: a histologic study in the canine mandible. Int J Oral Maxillofac Impl 1994;9:13-29.
8. Piattelli A, Scarano A, Quaranta M. High-precision, semi-automated, cost-effective system for producing thin sections of oral tissues containing dental implants. Biomaterials 1997;18:577-579.
9. Van Steenberghe D, Johansson C, Quirynen M, Molly L, Albrektsson T, Naert I. Bone augmentation by means of a stiff occlusive titanium barrier. A study in rabbits and humans. Clinical Oral Implants Research 2003;14:63-71.
10. Schmid J, Hämmene CHF, Olah A, Lang NP. Membrane permeability is unnecessary for guided generation of new bone. An experimental study in the rabbit. Clinical Oral Implants Research 1994;5:125-130.
11. Lundgren D, Lundgren AK, Sennérby L, Nyman S. Augmentation of intamembranous bone beyond the skeletal envelope using an occlusive titanium barrier. An experimental study in the rabbit. Clinical Oral Implants Research 1995;6:67-72.
12. Lundgren AK, Lundgren D, Taylor A. Influence of barrier occlusive-nas on guided bone augmentation. An experimental study in the rat. Clinical Oral Implants Research 1996;9:251-260.
13. Chierico A, Valentini R, Majzoub Z, Piattelli A, Scarano A, Okun L, Cordioli G. Electrically charged GTAM membranes stimulate osteogenesis in rabbit calvarial defects. Clinical Oral Implants Research 1999;10:415-424.